

Improvement effect of linseed oil on the activity of testes and physiological parameters in mice treated with Bicalutamide

Najdat Ali Al-kadhi^{1*}, Kasim Sakran Abass², Shaheen Ekram Jaafar¹

¹ Department of Medical Laboratory Techniques, Kirkuk Technical College, Northern Technical University, Iraq. ² Department of Pharmacology and Toxicology, College of Pharmacy, University of Kirkuk, Kirkuk, Iraq.

Correspondence: Najdat Ali Al-kadhi, Department of Medical Laboratory Techniques, Kirkuk Technical College, Northern Technical University, Iraq.

Email: al_kadhi2012@yahoo.com

ABSTRACT

The study aimed to explore the protective and improvement effect of linseed oil (LSO) on some physiological and histological parameters of male mice treated with Bicalutamide. The study carried out on 25 male mice, all treated orally once daily by gavage needle for 40 days. Animals were allocated randomly to 5 groups each included five at controlled conditions. First group served as control, 2nd group included castrated mice, 3rd group received LSO, 4th group received bicalutamide and 5th group received LSO+ bicalutamide. At the end of experiment, the blood collected and centrifuged to obtain serum and kept at deep freeze (-20 c) until biochemical tests carried out for detection of AST, ALT, ALP, urea, uric acid, creatinin and TSB by semi- auto chemistry analyzer. While testosterone, FSH, and LH were detected with automic fluorescence immune- assay. Histological study carried out after removal of testes to study the diameter and number of seminiferous tubules with number of leydig and sertoli cells. Results of the study recorded a significant elevation ($P<0.05$) in the concentration of AST, ALT, ALP, urea, uric acid, creatinin but significant reduction in the level of FSH, LH and testosterone in castrated and bicalutamide groups. While in the LSO and LSO+ bicalutamide groups not showed any remarkable differences ($P>0.05$) in their concentrations as compared control. However histological study concerned with the diameter of seminiferous tubules with number of leydig and sertoli cells showed significant reduction ($P<0.05$) in the bicalutamide group, while LSO and LSO + bicalutamide groups not recorded remarkable reduction ($P>0.05$). In conclusion supplementation of animals with LSO or co- administrated with bicalutamide can ameliorate and improves some physiological and testicular activities.

Keywords: Linseed oil, Bicalutamide, antiandrogen, hormones, male mice, hepatic enzymes.

Introduction

Linseed (*Linum usitatissimum*) belongs to the Linaceae family also known as flaxseed, found as seeds or oil forms, for its high nutritional quality recognized as a functional food [1]. Ancient Rome & Greece were used for the cure of different health complaints [2]. The plant is considered one of the richest plant sources for many trace elements and vitamins as vitamin B, E, and carotene [3]. Also, high levels of omega-3 PUFAs, specially α -

linoleic acid and high levels of vital oils [4], are positively associated with sperm morphology when highly consumed [5].

Essential oil of linseeds biologically precursor to Omega-3 fatty acids such as eicosapentaenoic acid (EPA) [6] are widely used in Asia and Europe for its potential anti-inflammatory effects attributed to the n-3 PUFAs contents that mediate the production of eicosanoids [7] which have antibacterial effects recorded by [8].

However, the plant have been found to have important beneficial effects for reduction concentration of bone biomarkers, plasma cholesterol and LDL-C [9], with high antioxidant capacity for inhibition of lipid peroxidation [10], and significantly decreases atherogenic risks [11]. While in vitro promotes apoptosis and showed inhibitory effects on growth [12]. Plant lignans such as secoisolaricinesinoldiglucoside (SDG) found in high concentration in Flaxseed which has been blocked cell proliferation and inhibition tumor growth, possibly by

Access this article online

Website: www.japer.in

E-ISSN: 2249-3379

How to cite this article: Najdat Ali Al-kadhi, Kasim Sakran Abass, Shaheen Ekram Jaafar. Improvement effect of linseed oil on the activity of testes and physiological parameters in mice treated with Bicalutamide. J Adv Pharm Educ Res. 2020;10(3):146-54. Source of Support: Nil, Conflict of Interest: Declared.

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-Non Commercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

modulating estrogen receptor or growth factor-dependent signaling in the experimental models ^[13] or the culture ^[14].

Male gonads secrete an important androgen (testosterone) which have sexual activities and beneficial effects for development and functions of the male reproductive system ^[15]. They are also responsible for prostate tumor growth ^[16]. 80-90% of prostate cancer is androgen-dependent ^[17].

Several anti-androgens including Bicalutamide can prevent and inhibit the influences of androgens by antagonist androgen receptors ^[18], while Finasteride inhibits 5-alpha reductase enzyme ^[19].

Bicalutamide chemically found as (2 RS)-4-cyano-3-(4-fluorophenylsulphenyl)-2-hydroxy-2-methyl-3-(trifluoromethyl)-propionanilide ^[20], an orally active non steroidal anti-androgen drug are commonly used in the treatment of prostate cancer but causes impairment of spermatogenesis and reduction of testicular weight ^[21] and behaves as pure anti-androgen that competitively blocks androgen receptors and inhibits both gene expression and cell growth ^[22]. It also exhibits androgen antagonistic activity ^[23], thereby inhibiting the negative feedback mechanism that regulates testosterone concentration ^[24].

The drug has prospective to interfere abnormally with the development as well as with the functions of animal and human reproductive system ^[25] that negatively affects the quality of life ^[16]. So different attempts were carried out to alleviate adverse effects or minimizing side effects of anti-androgenic drugs and improving the quality of life but without suitable results ^[26].

So this study was designed to investigate the ability of LSO supplementation for modulation and improvement of some physiological constituents and testicular activities of male mice treated with bicalutamide drug.

Materials and Methods

Materials:

Linseed oil prepared by (Emad company- Baghdad) purchased from a local pharmacy in Kirkuk city stored in a cool place and administered once in a day 1ml/kg B.W. orally by gavage needle ^[27]. While Bicalutamide drug is represented as bicalutamide 50 mg under trade name Casodex (AstraZeneca UK.), daily dose 1 tablet as manufacturer prescription.

Experimental design:

Twenty-five male albino Swiss mice, approximately two months old & weighing 26 ± 2 gm were housed (5 mice/cage) at controlled conditions were temperature 24 ± 2 °C and daily light / dark 14:10 hrs. Mice were fed rodent chow and tap water ad libitum. Animals of one group were castrated bilaterally under anesthesia while intact mice were used in all other groups.

Castration procedure:

A group of animals was castrated before 14 days of experimentation according to the technique described by ^[28], were animals injected general anesthesia with diazepam by I/P (10mg/ kg B.W.) followed 5 minutes later by ketamine (50 mg/kg B.W.). After initiation of anesthesia, the scrotal area was clipped then 70 % alcohol applied to disinfect the scrotum.

A small midline incision was made in the scrotal sac and each testis was delivered through the scrotal incision. Once exteriorized testis was removed after spermatic vessels were tied with 4.0 silk sutures. The incision was sutured an interrupted pattern to close the incision with dusting penicillin powder locally and I/m injection of penicillin G (2000 IU/kg B.W) for 3 days. Each castrated mice kept in an isolated cage till recovery.

Grouping and treatment:- Animals were allocated randomly to the following groups:-

- 1- The intact group that was treated with normal saline 0.1ml/mice served as a control group.
- 2- The castrated group that was treated with vehicle 0.1 ml/mice.
- 3- The intact group that was treated with linseed oil 1 ml/kg B.W/day. ^[27].
- 4- The intact group that was treated with Bicalutamide 0.8 mg/kg B.W.
- 5- Intact group treated with linseed oil 1ml/kg B.W + Bicalutamide 0.8 mg/kg B.W

All animals were treated orally once daily for 40 days by gavage needle. At the end of the experiment, blood was collected in jelly tubes by cardiac puncture, left for 10 minutes to clot, and centrifuged by Centrifuge (Centrion UK) for 5000 rpm for 10 minutes. Serum was kept in Eppendorf tubes and stored at deep freeze (-20°C) until the biochemical analysis was carried out.

Biochemical assays:- levels of hepatic enzymes in the serum, aspartate aminotransferase (AST), alanine aminotransferase (ALT), and alkaline phosphatase (ALP). Renal function tests included urea, uric acid, creatinin, and total serum bilirubin concentration. All biochemical parameters are estimated with commercially available kits AGAPPE (AGAPPE diagnostic Switzerland GmbH) and assayed by Semi-Auto Chemistry Analyzer (Mindray BA-88A) at 540 nm as prescribed by the manufacturer information.

Hormones in the serum of male mice: testosterone, FSH, and LH levels were assessed by commercial kits, and detected with Automatic fluorescence Immuno-Assay (AFIAS 6) as depicted by manufacturer information.

For histological studies, the animals were killed by decapitation under basal conditions in a longitudinal fashion. Testes, from each animal, were weighted, immersed in 10% buffered formalin for routinely processed in a tissue processor, and embedded in paraffin. Sections 5 µm were cut by microtome and stained with hematoxylin and eosin stain ^[29]. Examination of prepared slides

performed by a light microscope with a camera (Optica, Italy). The diameter of seminiferous tubules was carried out by ocular micrometer after calibration with micrometer stage ^[30], and also, some seminiferous tubules, Leydig cells and Sertoli cells were determined ^[31].

Statistical Analysis:

Conventional statistical methods were used to calculate means and standard errors. Analysis of variance (ANOVA) was applied to test for any significant differences ($P < 0.05$). All statistics were carried out used SigmaPlot (version 12).

Results:

The results of the study recorded alterations of some biochemical parameters concerned with hepatic enzymes and TSB were detected in the serum of treated animals as shown in table 1, were a group of mice treated with the anti-androgenic drug (Bicalutamide) 0.8mg/kg B.W. daily for 40 days recorded a significant elevation ($p < 0.05$) in the concentration of AST (229.4 ± 4.4), ALT (28.4 ± 2.76), ALP (488.8 ± 12.02) and TSB (0.68 ± 0.03) as compared to the control and other treated groups, but castrated group recorded a significant increase ($p < 0.05$) in the level of ALP (386.4 ± 2.85) as compared to the control group. While LSO and LSO + Bicalutamide treated groups recorded non-significant differences ($p > 0.05$) in the activities of hepatic enzymes and TSB as compared to the control group, although LSO treated group recorded significant decrement ($p < 0.05$) in the level of AST (125.8 ± 3.29) as compared to the control group.

On other hand, the other biochemical parameters: urea, uric acid, and creatinine concentration revealed also in table 1, were recorded remarkable elevation ($p < 0.05$) in the group of animals treated with bicalutamide (55 ± 2.64), (362.2 ± 4.42) and (0.62 ± 0.03) respectively as compared control and other treated groups. Whereas the Castrated group recorded a significant increment ($p < 0.05$) in the activities of urea (45.4 ± 2.52), uric acid (209 ± 2.64), and creatinin (0.52 ± 0.03) as compared to the control, LSO and LSO + bicalutamide treated

groups. While LSO and LSO + bicalutamide treated groups recorded a non-significant decrease ($p > 0.05$) in the concentration of estimated biochemical parameters as compared to the control group.

Table 2 shows the results of the hormonal activities, FSH, LH, and testosterone in the serum of control and treated animals were recorded in the castrated and bicalutamide treated groups a significant decline ($p < 0.05$) in the concentration of FSH (0.19 ± 0.01) (0.28 ± 0.02), LH (0.14 ± 0.02) (0.34 ± 0.02) and testosterone (0.66 ± 0.05) (0.72 ± 0.03) respectively as compared to the control other and treated groups.

While LSO treated group recorded a significant increment ($p < 0.05$) in the level of testosterone (2.50 ± 0.1) and non significant increment ($p < 0.05$) in the level of FSH (0.66 ± 0.05) and LH (0.82 ± 0.03) as compared to the control group. Whereas, LSO+ bicalutamide treated group didn't recorded significant decrease ($p > 0.05$) in the activities of FSH (0.50 ± 0.03), LH (0.65 ± 0.03) and testosterone (1.65 ± 0.02) as compared control and LSO treated groups.

Histological parameters of testes shown in the table, 3 and figure, 1 revealed that the treatment of animals with bicalutamide recorded a significant reduction ($p < 0.05$) in the diameter of seminiferous tubules (258 ± 2.49), number of Leydig cells (27.9 ± 1.43) and Sertoli cells (20.6 ± 0.87) as compared to the control and other treated groups. While LSO treated group not recorded significant increment ($p > 0.05$) in the diameter of seminiferous tubules (289.4 ± 5.28) and number of seminiferous tubules (22.1 ± 1.16), Leydig cells (58.6 ± 1.8) and Sertoli cells (31.8 ± 1.06) as compared to the control group, but the increment was in above parameters significantly ($p < 0.05$) when compared with the bicalutamide treated group.

On other hand, co-treatment of animals with LSO + bicalutamide recorded a remarkable increment ($p < 0.05$) in the number of Leydig cells (39.8 ± 1.31) and Sertoli cells (25.8 ± 0.73) but non-significantly ($p > 0.05$) in the diameter and number of seminiferous tubules (278.4 ± 2.5), (21.2 ± 0.8) respectively as compared to the bicalutamide treated group. However, there were no significant differences ($p > 0.05$) in the number of seminiferous tubules between all treated groups.

Table 1: Effect of Castration, Bicalutamide, LSO, and LSO+Bicalutamide treatment on the concentration of some biochemical parameters ($M \pm SE$) in the serum of male mice.

Groups Parameters	Control	Castrated	LSO	Bicalutamide	LSO+ Bic.
AST	177.20 ± 2.28	180.6 ± 3.05	125.8 ± 3.29	229.4 ± 4.40	171.8 ± 4.69
U/L	a	a	b	c	a
ALT	15.20 ± 1.06	24.6 ± 1.72	16.8 ± 0.96	28.2 ± 2.76	22.4 ± 1.03
U/L	a	bc	a	c	ab
ALP	338.2 ± 2.69	386.4 ± 2.85	320.8 ± 3.0	488.8 ± 12.02	345 ± 4.62
U/L	a	b	a	c	a
TSB	0.34 ± 0.02	0.42 ± 0.03	0.26 ± 0.02	0.68 ± 0.03	0.44 ± 0.05
mg/dl	a	a	ab	c	a
Urea	33.8 ± 1.59	45.4 ± 2.52	25.8 ± 1.49	55.0 ± 2.64	33.0 ± 1.84
mg/L	a	b	a	c	a
Uric acid	181.8 ± 2.65	209.0 ± 2.64	142.0 ± 2.62	362.2 ± 4.42	177.4 ± 2.82
mmol/L	a	b	c	d	a
Creatinin	0.36 ± 0.02	0.52 ± 0.03	0.32 ± 0.03	0.62 ± 0.03	0.36 ± 0.04
mg/dl	a	b	a	b	a

a, b, c: small letters refer to represent significant differences ($p < 0.05$) between groups at the horizontal arrows.

Table 2: Effect of Castration, Bicalutamide, LSO and LSO+Bicalutamide treatment on the concentration of some hormones (M±SE) in the serum of male mice

Groups	Parameters	Control	Castrated	LSO	Bicalutamide	LSO+Bic.
	FSH mlu/ml	0.53 ± 0.03 a	0.19 ± 0.01 b	0.66 ± 0.05 c	0.28 ± 0.02 b	0.50 ± 0.03 ac
	LH mlu/ml	0.72 ± 0.03 a	0.14 ± 0.02 b	0.82 ± 0.03 a	0.34 ± 0.02 c	0.65 ± 0.03 a
	Testosterone ng/ml	1.74 ± 0.01 a	0.66 ± 0.05 b	2.50 ± 0.1 c	0.72 ± 0.03 b	1.65 ± 0.02 a

a, b, c: small letters refer to represent significant differences (p<0.05) between groups at the horizontal arrows.

Table 3: Effect of Castration, Bicalutamide, LSO, and LSO+bicalutamide treatment on the diameter and number of seminiferous tubules, Leydig, and Sertoli cells (M±SE) in the testes of male mice.

Groups	Parameters	Control	LSO	Bicalutamide	LSO + Bic
	Seminifer tubules(μm)	298.8 ± 22.8 a	284.4 ± 5.28 a	258.6 ± 2.49 B	278.4 ± 2.5 ab
	Seminifer tubules(No.)	20.6 ± 1.20 a	22.1 ± 1.16 a	18.8 ± 0.51 A	21.2 ± 0.80 a
	Leydig cells (No.)	53.2 ± 2.15 a	58.6 ± 1.8 a	27.6 ± 1.43 B	39.8 ± 1.31 C
	Sertoli cells (No.)	30.0 ± 1.30 a	31.8 ± 1.06 a	20.6 ± 0.87 B	25.8 ± 0.73 C

a, b, c: small letters refer to represent significant differences (p<0.05) between groups at the horizontal arrows.

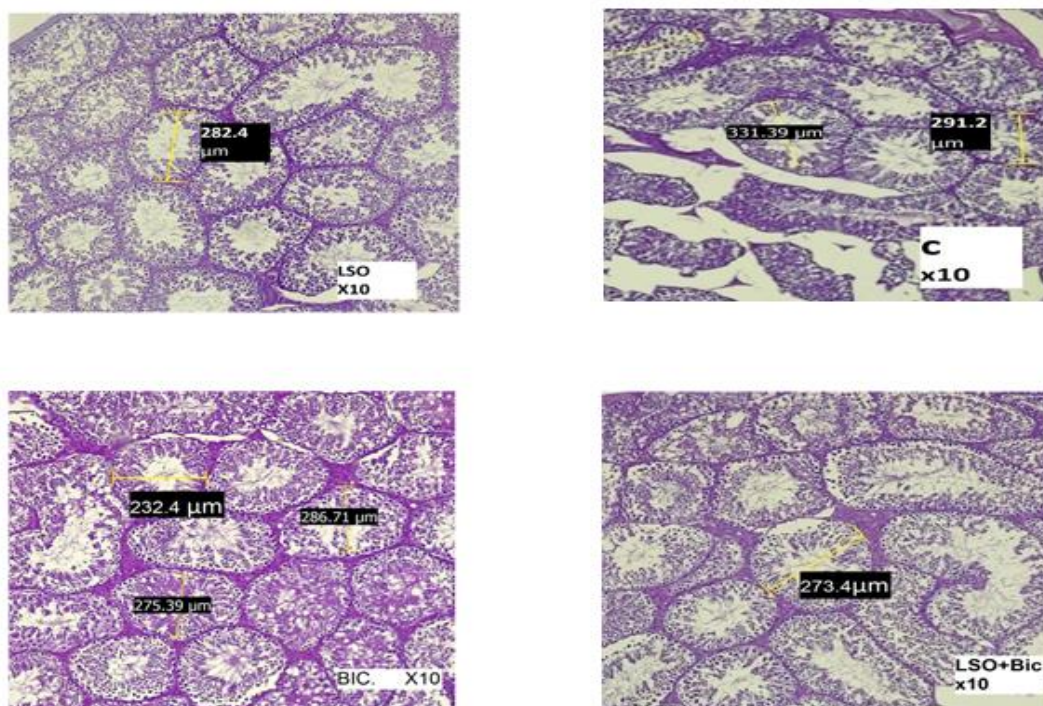


Figure 1: Shows effects of bicalutamide, LSO, and LSO+bic. Treatment following hematoxylin and eosin staining(x10) on the measurement of seminiferous diameter and number with the number of leydig and sertoli cells in the tissues of testes of male mice.

C: control group, Bic: bicalutamide treated group, LSO: Linseed oil treated group, and LSO+Bic: group co-treated LSO with bicalutamide.

Discussion:

Treatment of animals for forty days with the anti-androgenic drug(bicalutamide) associated with several side effects that have

a significant impact on the quality of life, as revealed from results of the current study were recorded many adverse effects concerned firstly with the biochemical parameters as shown in (table 1) were concentration of AST, ALT and ALP enzymes in

addition to TSB and renal function test parameters, urea, uric acid, and creatinin were recorded a significant elevation ($p < 0.05$) as compared to the control and other treated groups. Suggesting that the treatment of animals for a long period with bicalutamide may cause intoxication or oxidative stress which could play an important role in hepato-renal damage. Damages for renal and hepatic cells reflect an elevation in the hepatic enzymes and renal function test parameters in the serum of intoxicated or oxidative stress animals [32].

Results of the current study were compatible with the results of many studies they recorded an increase of hepatic enzymes significantly in rats administrated bicalutamide repeatedly 25 mg/kg B.W. [18], the same results recorded also in rats exposed to diazinon an organophosphorus insecticide [33]. Increased hepatic enzymes concentration in the serum concerned common adverse effects for bicalutamide administration, so regular liver function tests during treatment must be advised [34].

The impairment of renal function test parameters in the current study as shown in table 1, confirmed with the elevation of urea, uric acid, and creatinin concentration in the serum of treated group were also documented by several studies. The exposure of rats to lead acetate induced damage for renal tissues leads to an elevation of urea, uric acid, and creatinin level [35]. However, animals exposed to diazinon recorded a significant increase in the parameters of the renal function test [5].

Therefore, dietary supplements may be beneficial for animals undergoing anti-androgenic therapy, so the results of this study showed LSO supplementation attenuated the extensive alterations of the biochemical parameters in the bicalutamide treated group. The results of the study are not recorded significant differences ($p > 0.05$) in all biochemical parameters as compared to the control group, moreover, these results were more clear in LSO treated group (table 1). Administration of FXO to rats treated with lead acetate improved levels of urea, uric acid, and creatinin in the serum of animals [35]. Comparative study about Improvement and protective effects between liv.52 and silymarine supplementation to rats induced hepatic damage showed liv.52 was more effective than silymarine [18]. When a group of mice is exposed to 5 Gy of gamma radiation, the induced elevation of hepatic enzymes in which their concentrations declined and normalized in pretreated with FXO for 15 consecutive days during the exposure period [36].

Castration is also one of the methods for deprivation of androgens concentration in the serum and one of the treatment methods for prostate cancer but it is currently limited in developed countries. But this procedure included many deleterious side effects similar to the anti-androgenic drugs particularly that concerned with the biochemical and hormonal criteria (table 1 and 2). So almost the same results were recorded in the castrated group as found in the bicalutamide treated group concerned with hepatic and renal function test parameters, suggesting the castration may cause some damages for tissues around incision site or may generate oxidative stress from castration operation. The destruction of

tissues with oxidative stress increases the release of transaminases enzymes and elevates their concentrations in the serum. The generation of ROS and depletion of the thiol system mediates hepatic and renal damages increase the concentration of hepatic & renal function test parameters [37]. Results of the castration on primate rhesus monkey recorded a significant increment in the levels of ALT, ALP, urea, and bilirubin [38], while the rats which not recorded significant differences, because these rats replaced with testosterone hormone after castration [39].

Table 2, showed significant decline ($p < 0.05$) in the activities of FSH, LH and testosterone hormones in the serum of castrated and bicalutamide treated animals, where the results were in full agreement with the other studies, suggesting that the regulation of hormones can be interrupted and disrupted at hypothalamo-pituitary level particularly FSH and LH secretion.

Different conformational changes in adult male rats produced by the anti-androgens that competitively block androgenic receptors, therefore, avoided participation of testosterone in the cellular process [40]. Results of the study on the primate rhesus monkey castration recorded a significant decline in testosterone hormone while an increment in the levels of ALT, ALP, urea, and bilirubin in the serum of animals [38].

Cellular mitosis and meiosis of germ cells with the development of the reproductive system in male rats were achieved essentially via interaction of testosterone with androgen receptors on target cells [41]. Treatment with bicalutamide might adversely affects Leydig cells that cause a significant decrease in testosterone synthesis and release [42]. While a slight decline was recorded in the study of [43]. Decrement of testosterone level in rats recorded after oral administration of bicalutamide, these results also achieved in human [44], While not found any changes in the study of [45]. Suppression of anterior pituitary feedback by testosterone in rats was one of the deleterious adverse effects of bicalutamide treatment, which causes a reduction of testicular weight and related changes in testicular tissues [46]. Moreover, exposure of human populations to the anti-androgenic and estrogenic chemicals in the environment can interfere with the level of endogenous sex hormones [47].

Nonetheless, LSO treatment alone or co-treatment with the bicalutamide showed a significant increase ($p < 0.05$) in the studied hormones as revealed in (Table 2). From our results, we found that supplementation of LSO elevated concentration of FSH, LH, and testosterone suggesting the relationship between LSO and the reproductive endocrine system. Furthermore, LSO rich in many active components that may have ameliorative and enhancement effects on testicular tissue or may have a direct role for the regulation of hormones at the hypothalamo-pituitary gonads axis.

Moreover, PUSFAs in the LSO are direct precursors for a large group of compounds that physiologically active [48]. However, saponins may stimulate endogenous testosterone probably by increases the level of LH or may bound to the enzymes involved in steroidogenesis in male rats supplemented with phoenix

doctylifera extract ^[49]. On other hand, supplementation of rats with *Nigella sativa* oil recorded a significant increase in the concentration of FSH and LH may be as a result of direct stimulatory effects of oil upon the hypothalamus to increase GnRH secretion ^[50]. Similar effects were also observed on the level of testosterone when supplemented male rats with the aqueous extract of *Rauvolfia* ^[51].

Table 3, and figure 1, showed histopathological observation deformities at the testicular photometric parameters of bicalutamide treated animals, with serious damages within testis cells and diameters of seminiferous tubules. These results suggest the significant disturbances in the activities of FSH, LH, and testosterone hormones, therefore physiological influences may be interrupted and disrupted at the tissues of the testicular level. Moreover, bicalutamide competitively binds with androgen receptors that inhibit the stimulatory effects of hormones, therefore enhances apoptosis the cellular structures of the primary and secondary male reproductive organs. When a group of rats exposed to bicalutamide ^[52] and other groups of rats to flutamide ^[53] accelerated the apoptotic process in cells associated with the enhances the expression and activation of caspases-3 and caspases-6 enzymes were both are essential elements in the apoptosis of cells. Reduction of testicular weight with significant alterations in the tissues of testis after treatment of rats with bicalutamide were recorded, and adverse effects of bicalutamide may be attributed to the adverse effects the drug ^[54], with the suppression of testosterone on the anterior pituitary feed-back ^[55]. In males, secretion of GnRH from the hypothalamus stimulates Leydig cells within interstitial cells of the testes to synthesize and release of testosterone hormones, while FSH stimulates development and differentiation of Sertoli cells and give rise to the processes of spermatogenesis ^[56]. Retardation growth and development of germ cells in the rat testes were investigated due to the remarked decrease in the level of testosterone when treated rats with bicalutamide ^[57]. Similar effects were also recorded when treated rats with flutamide ^[53]. Also, Khursheed, et al 2011, recorded a significant reduction in the diameter and germinal layer thicknesses of the seminiferous tubules in the bicalutamide treated group.

Dietary supplements have remarkable beneficial on subjects undergoing anti-androgenic therapy, so the results of LSO supplementation in this study, revealed the alleviation of the extensive changes in the histopathological and architecture of testicular parameters in animals treated with bicalutamide as shown in (Table 3, and figure 1). These results suggest that the treatment of animals with LSO may act directly or indirectly on the sectional functions of the pituitary gland that stimulates testicular cells which control spermatogenesis and steroidogenesis. Whereas reproductive functions are androgen-dependent, therefore any changes in androgen secretion would reflect and explain alterations in the number of Leydig cells with their functions.

Protective and therapeutic properties of LSO are mostly attributed to the high contents and richness of LSO for many viable USFAs, α -linolenic acid also high contents of lignan precursor responsible for their antioxidant activities ^[58]. Oxidative damages in testes of rats induced by the cadmium were attenuated & improved by the FXO administration ^[59]. Improvement of histopathological changes afforded by FXO may belong to the beneficial effects of PUSFAs that may inhibit lipid peroxidation and subsequently alters membrane fluidity ^[60]. In addition to the high contents of FXO for oleic acid, monounsaturated fatty acids that decrease the susceptibility of the testicular tissue to lipid peroxidation which responsible for integrity defects of seminiferous tubules ^[61].

Testicular toxicity in male rats induced by sodium valerate was ameliorated and protected by the supplementation with LSO was more successful than *Nigella sativa* and Celery oils ^[62]. Moreover, treatment of male rats with lead acetate recorded degenerative alterations in testicular tissue were rescued significantly after treatment with FXO. Cured effect of FXO postulated to the enhancement influences of the plant on the healthy seminiferous tubules filled with different germ cells including Leydig and Sertoli cells ^[8].

Conclusion: Anti-androgenic drug bicalutamide have several adverse effects and most of them can modulated and ameliorated particularly biochemical, hormonal, and testicular criteria when LSO administrated alone or co-treated with bicalutamide.

Conflict of Interests

The authors of this paper declare that he has no financial or personal relationships with individuals or organizations that would unacceptably bias the content of this paper and therefore declare that there is no conflict of interests.

Acknowledgments

This research project is funded by itself. The author wishes to thanks the Dean of College of Veterinary Medicine at Kirkuk University who contributed to the study and the animal health assistants and laboratory technicians for their expert work in collecting the samples and for their useful assistance

References

1. Adolphe J.L., Whiting S.J., Juurlink B.H., Thorpe L.U., Alcorn J. Health effects with consumption of the flax lignan secosolariciresionaldiglucoside. *Br. J. Nutr.* 2010; 103 : 929-938.
2. Goyal A., Sharma V., Upadhyay N., Gell S., Sihag M. Flax, and flaxseed oil: an ancient medicine and modern functional food. *J. Food Sci. Technol.* 2014; 51: 1633-1653

3. Hana RS. and Sead N. Alteration in oxidants, antioxidants, and cytokines levels in blood of malathion exposed human and animal groups and the effect of flaxseed oil in alleviating malathion toxic effects. *Euro J BiotechnolBiosci.* 2013; 1(1):8-19.
4. Barcelo- Coblijn G, Murphy EJ Alpha-linolenic acid and its conversion to longer chain n-3 fatty acids: benefits for human health and a role in maintaining tissue n-3 fatty acid levels. *Prog in lipid Res.* 2009; 48: 355-374.
5. AL-Attar, A.M., Abu Zaid, I.M., Effect of tea (*Camellia sinensis*) and olive (*Olea Europaea L.*) leave extracts on male mice exposed to diazinon. *Int. Bio. Med. Res.* 2013; 1-6.
6. Prasad, K. Flaxseed: A source of hypocholesterolemic and antiatherogenic agents. *Drug News and perspectives.* 2000; 13(2): 99-104.
7. Cohen, S. L., Moore, A. M., Ward, W. E. Flaxseed oil and inflammation-associated bone abnormalities in interleukin-10 knockout mice. *Journal of Nutritional Biochemistry.* 2005; 16 (6): 368-374.
8. Abdel-Monem, A., Dkhil, M., Al-Quraishy, S. Protective role of flaxseed oil against lead acetate induced oxidative stress in testes of adult rats. *Afr. J. Biotech.* 2010; 9: 7216-7223.
9. Boulbaroud, S., El-Hessni, A., Azzaoui,F-Z., Mesfioui A. Sesame seed oil, and flaxseed oil affect plasma lipid levels and biomarkers of bone metabolism in ovariectomized Wistar rats. *Biology and Medicine*,2010. 4 (3): 102-110.
10. Sargi CS, Silva CB, Santos CMH, Montanher FP, Boeing SJ, Junior SOO, Souza EN, Visentainer VJ Antioxidant capacity, and chemical composition in seeds rich in omega-3: chia, flax, and perilla. *Food SciTechnol Campinas.* 2013; 33(3):541-548
11. Molfino A., Gioia G., Rossi Fanelli F., Muscaritoli M. The role for dietary omega-3 fatty acids supplementation in older adults. *Nutrients.* 2014; 6: 4058-4073.
12. Mansara P.P., Deshpande R.A., Vaidya M.M., Kaul-Ghanekar R. Differential ratios of omega fatty acids (AA/ EPA + DHA) modulate growth, lipid peroxidation and expression of tumor regulatory MARBPs in breast cancer cells in high and low estrogen environments. *Nutr. Cancer.* 2015; 67: 1001-1009.
13. Chen J., Power K.A., Mann J., Chen A., Thompson L.U. Flaxed alone or in combination with tamoxifen inhibits MCF-7 breast tumor growth in ovariectomized athymic mice with high circulating levels of estrogen. *Exp. Biol. Med.* 2007; 232: 1071-1080.
14. Sagar J.K., Chen J., Corey P., Thompson L.U. The effect of secoisolariciresinol diglucoside and flaxseed oil, alone and in combination, on MCF-7 tumor growth and signaling pathways. *Nutr. Cancer.* 2010;62, p:533- 542
15. Heinlein, C.A., Chang, C. Androgen receptor in prostate cancer. *Endocr. Rev.* 2004, 25, 276-308.
16. Andrea Dueregger, Isabel Heidegger, Philipp Ofer, Bernhard Perktold, Reinhold Ramoner, Helmut Klocker, and Iris E. Eder. The use of Dietary Supplements to Alleviate Androgen Deprivation Therapy Side Effects during Prostate Cancer treatment. *Nutrients.* 2014; 6, 4491-451.
17. Denis, L.J. The role of active treatment in early prostate cancer. *Radiother. Oncol.* 2000, 57, 251-258.
18. Fulzele V.B., Shedage A.T., Anton Smith A., Gaikwad T.V., Kirtane S.R. Comparative hepatoprotective activity of liv-52 and silymarine against hepatotoxicity induced by antiandrogen- bicalutamide in rats. 2012; 4, suppl. 3, 211-213.
19. Monk JP, Halabi S, Picus J, Hussain A, Philips G, Kaplan E, Ahles T, Gu L, Vogelzang N, Kelly WK, Small EJ. Efficacy of peripheral androgen blockade in prostate cancer patients with biochemical failure after definitive local therapy. *Cancer.* 2012; 1: 4139-47.
20. Fradet, Y. Bicalutamide (Casodex ®) in the treatment of prostate cancer. *Expert review of anticancer therapy.* 2004; 4, 37-48.
21. Ahmed E. Abdel Moniem, Mohamed A. Dkhil, Saleh L-Quraishy. Protective role of flaxseed oil against lead acetate induced oxidative stress in testes of adult rats. *African Journal of Biotechnology.* 2010; 9 (42): 7216-7223.
22. Khursheed A, Minhas LA, Niaz WA. Histomorphometric study of effects of bicalutamide on spermatogenesis in male rats. *Pak. Armed Forces Med. J.* 2011; 61(3): 325-9.
23. Terouanne, B., Tahiri, B., Georget, V., Belon, C., Poujol, N., Avances, C., Orio JR, f., Balaguer, p., Sultan, C. A. Stable prostatic bioluminescent cell line to investigate androgen and antiandrogen effects. *Molecular and cellular endocrinology.* 2000; 160, 39-49.
24. Mcleod, D.G., Iversen, P. Gynecomastia in patients with prostate cancer: a review of treatment options. *Urology.* 2000; 56, 713-720.
25. Sharpe, R. M. Pathways of endocrine disruption during male sexual differentiation and masculinization. *Best Pract Res Clin Endocrinol Metab.* 2006; 20, 91-110.
26. Grossmann, M., Zajac, J.D. Androgen deprivation therapy in men with prostate cancer: How should the side effects be monitored and treated? *Clinical. Endocrinology.* 2011, 74, 289-293.
27. Kaithwas G., Majumder DK. In vitro antioxidant and in vivo antidiabeticantihyperlipidemic activity of linseed oil against streptozotocin-induced toxicity in albino rats. *Eur. J. Lipid Sci. Tech.* 2012; 114:12-45.
28. Watcho P, Wankeu-Nya M, Ngudefack TB, Tapondjou L, Teponno R, Kamanyi A. Pro- sexual effects of *Dracaena Arborea* (wild) link (*Dracaenaceae*) in sexually

- experienced male rats, *Pharmacology online*. 2007; 1:400-19.
29. Luna L.G. Manual of histologic staining methods of the armed forces institute of pathology. 3rd ed. Me Graw- Hill Book company. Newyork. (1968).
30. Galigher A.E., Kozolof E.N. Essential of practical microtechnique. Lea and Fabigher. Philadel. Lst.ed. 1964.
31. Fereshteh Mehaïen and FeraidoonNegahdar. Morphometric evaluation of seminiferous tubules in aged mice testes after melatonin administration. *Cell journal*. 2011. 13(1). P: 1-4.
32. Azheen S. Abdulrahman, Inaam A. Mustafa. Impact of bicalutamide, an anti-androgen on rat testis. *Zanco Journal of pure and applied sciences*. 2019, 31(2). P:89-100.
33. EL-Demerdash, F.M., Nasr, H.M., Antioxidant effect of selenium on lipid peroxidation, hyperlipidemia, and biochemical parameters in rats exposed to diazinon. *Trace Elem. Med. Biol.* 2014, 28, 89-93.
34. Chianakwalm C.I. et al, A case of male breast cancer in association with bicalutamide-induced gynecomastia, *The Breast*. 2005; 14(2):163-164.
35. Ahmed E.AbdelMoneim, and Saleh AL-Quraishy. The protective effect of flaxseed oil on lead acetate-induced renal toxicity in rats. *Journal of Hazardous Materials*. 2011; Vol.194, PP: 250-255.
36. Bhatia, A.L., Avadhesh Sharma, ShikhaPatni, AntimLata Sharma. Prophylactic effect of flaxseed oil against radiation-induced. *PhytotherapyResearsch*. 2007; 21, (9):119-225.
37. Tabassum H, Suhel parvez, Hasibur Rehman, Basu Dev, Banerjee, Detlef Siemen, Sheikh Raisuddin. Nephrotoxicity and its prevention by taurine in tamoxifen induced oxidative stress in mice. *Human and experimental toxicology*. 2007; 26:509-18.
38. Nagarajan P. Arindkar S., Singh S., Majumdar S. Effect of long-term castration on serum biochemistry in rhesus monkeys. *Journal of Medical Primatology*. 2013. 42, (3):128-136.
39. Hassan A. A. Effect of Castration on Some Physiological Aspect in Rats: Effect of Testosterone Hormone. *J.Edu.&Sci*. 2010. 23(3), : 28-39.
40. AtikaKhursid, Shabnam Hamid, WaqarAzimNiaz. Effect of bicalutamide; an antiandrogen, on testicular weight in adult rats. *Pak. Armed Forces Med. J.* 2014; 64 (2): 204-07
41. Brinkmann AO, Block LJ, De Ruiter PE, Doesburg P, Steketee K, Berrevotes CA. Mechanisms of androgen receptor activation and function. *J. Steroid Biochem. Mol. Biol.* 1999; 69 (1-6): 307-13.
42. Chiao, Y.-C., Cho, W.-L., Wang, P. S. Inhibition of testosterone production by propylthiouracil in rat Leydig cells. *Biology of reproduction*. 2002; 67, 416-422.
43. Han, J.-H., Shin, M.-S., Lee, J.-M., Kim, T.-W., Jin, J.-J., KO, I.-G., Kim, C.-J., Kim, M., ROH, J. H. Long-term chemical castration induces depressive symptoms by suppressing serotonin expression in rats. *Animals Cells and systems*. 2018; 22, 29-36.
44. Hashimoto, K., Masumori, N., Hashimoto, J., Takayangi, A., Fukuta, F., Tsukamoto, T. Serum testosterone level to predict the efficacy of sequential use of antiandrogens as second-line treatment following androgen deprivation monotherapy in patients with castration-resistant prostate cancer. *Japanese journal of clinical oncology*. 2010, 41, 405-410.
45. Morgante, E., Gradini, R., Realacci, M., Sale, P., Deramo, G., Perrone, G., Cardillo, M., Petrangeli, E., Russo, M., Di Silverio, F. Effects of long –term treatment with antiandrogenbicalutamide on human testis: an ultrastructural and morphometric study. *Histopathology*. 2001; 38, 195-201.
46. Wason,S., Pohlmeier-Esch, G., Pallen, C., Palazzi, X., Espuna, G., Bars,R. 17 α -methyltestosterone: 28-day oral toxicity study in the rat based on the ‘ enhanced OECD test guideline 407’ to detect endocrine effects. *Toxicology*. 2003; 192, 119-137.
47. Benson T. Akingbemi, Matthew P. Hardy. Oestrogenic and antiandrogenic chemicals in the environment: effects on male reproductive health. *Annals of Medicine*. 2001, Volume 33, Issue 6. pp: 391-403.
48. Needleman P, Turk J, Jakschik BA, Morrison AR, Lefkowitz JB: Arachidonic acid metabolism. *Ann Rev Biochem*.1986; 55:69-102.
49. Abedi A, Parviz M, Karimian MS, Rodsari SRH. Aphrodisiac activity of aqueous extract of *Phoenix dactylifera* pollen in male rats. *Adv Sex Med*. 2013; 3: 28-34.
50. JumaFT., Hayfaa MA.: The effects of *Nigella sativa* oil administration on some physiological and histological values of reproductive aspects of rats. *Iraqi J. Vet. Med*. 2011; 35:52-60.
51. Lembe MD, Koloko LB, Bend FE, Domkam J, Oundoum OPC, Njila NM, Moundipa P, Dimo T, Gonzales GF. Fertility enhancing effects of aqueous extract of *Rauvolfia vomitoria* on reproductive functions of male rats. *J ExplIntegr Med*. 2014. Vol.4 Suppl(1):43-9
52. Saikumar, P. and Venkatachalam, M. A. Apoptosis, and cell death. *Basic Concepts of Molecular Pathology*. Springer; 2009.
53. Omezzine, A., Chater, S., Mauduit, C., Florin, A., Tabone, E., Chuzel, F., Bars, R. &Benahmed, M. Long-term apoptotic cell death process with increased expression and activation of caspase-3 and-6 in adult rat germ cells exposed in utero to flutamide. *Endocrinology*. 2003; 144, 648-661.

54. Khursheed, A., Hamid, S., Niaz, W. A. Effect of bicalutamide; an antiandrogen on testicular weight in adult rats. *Pak Armed Forces Med j.* 2014; 64, 204-207.
55. Kennel, P., Pallen, C., Barale- Thomas, E., Espuna, G., Bars, R. Tamoxifen: 28-day oral toxicity study in the rat based on the Enhanced OECD test Guideline 407 to detect endocrine effects. *Archives of toxicology.* 2003; 77, 487-499.
56. Martin GB, and Walkden-Brown SW: Nutritional influences on reproduction in mature male sheep and goats. *J. Reprod. Fertil.* 1995; Suppl: 49: 437-499.
57. Vidal, J. D., and Whitney, K. M. Morphologic manifestations of testicular and epididymal toxicity. *Spermatogenesis.* 2014. 4(2): 123-128.
58. Ethan Basch, Mphil, Steve Bent, Jeffrey Collins, Cynthia Dacey. Flax and Flaxseed Oil (*Linum usitatissimum*): A Review by the Natural Standard Research Collaboration. *Journal of the Society for Integrative.* 2007. Vol. 5, No 3, pp: 92-105.
59. Karaca S, Eraslan G. The effects of flaxseed oil on cadmium-induced oxidative stress in rats. *Boil Trace Elem. Res.* 2013; 155:423-430
60. Best CA, Cluette- Brown JE, Teruya A, Laposata M. Red blood cell fatty acid ethyl esters: a significant component of fatty acid ethyl esters in the blood. *J. Lipid Res.* 2003; 44: 612-620.
61. Bourre JM, Dumont O., Durand G. Effect of dietary oleic acid: oleic acid is conditionally essential for some organs. *Reprod. Nutr. Dev.* 2004; 44 (4): 371-380.
62. Hala, Maram A., Wahba, A. Protective Effect of Nigella Sativa, Linseed, and Celery Oils against Testicular Toxicity Induced by Sodium Valproate in Male Rats. *J. of American Science.* 2011; 7 (5); P: 687-693.