

# Microbiological evaluation of Sausages retailed for sale in Al Ramadi, Iraq

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## ABSTRACT

Meat products are an excellent source of a wide variety of nutrients, high quality proteins, vitamins and certain minerals. At the same time, they contain an abundance of all nutrients required for the growth and multiplication of most microorganisms. The purpose of the current work was to make a microbiological evaluation of sausages retailed for sale in Al Ramadi, Iraq. To achieve this goal, a total of 100 fresh chicken and beef sausage samples (50 / each) were randomly collected from different shops. The obtained results clarified that the mean values of aerobic bacterial, coliforms, *Staphylococcus aureus*, *Escherichia coli*, *Salmonella spp.* and *Shigella spp.* counts for chicken sausages were  $3.65 \times 10^5$ ,  $5.35 \times 10^4$ ,  $0.4 \times 10^2$ ,  $1.85 \times 10^3$ ,  $0.04 \times 10^2$  and  $0.005 \times 10^2$  cfu/g, respectively. On the other hand, they were  $2.6 \times 10^6$ ,  $1.9 \times 10^6$ ,  $0.85 \times 10^2$ ,  $2.5 \times 10^3$ ,  $0.085 \times 10^2$  and  $0.015 \times 10^2$  cfu/g for beef sausages, respectively. In conclusion, it was clear that the mean values of counts for studied microbiological criteria were higher in beef sausages compared to chicken sausages reflecting a contaminative environment during processing, handling, storage and retailing of beef sausages as compared to that of chicken sausages. Finally, it was proved that contaminated food had been shown to be a critical link in transmitting pathogens to humans resulting in numerous cases of diseases. Therefore, great emphasis should be placed on the microbiological aspects of sausages and on searching for alternative mechanisms to reduce both natural and cross contamination, thus avoiding major public health problems; so, it is important to adopt hazard analysis and critical control point principles in processing and handling of sausage to achieve pathogen free products.

**Keywords:** Sausages, Microbiological evaluation, Al Ramadi, Iraq.

## Introduction

Technology development in poultry meat processing and handling has given consumers a much greater choice over the food they can buy, so meat hygiene can comprise every step of processing from the health of the life bird to the distribution of the final product. It prevents using of harmful ingredients in manufacturing of poultry meat products as well as prevents the sale of contaminated or unwholesome meat.

Sausage is considered one of most beef and poultry meat suitable for human consumption. The English word "Sausage" is derived from the Latin word "Salsus", which means salted meat or salt preserved meat. Sausages are considered in general among the most widely consumed foodstuff by humans. It is imported from several countries and is also produced locally. They are crushed thoroughly with water (ice), table

salt, sodium nitrate or potassium by adding one or more permitted nutritional additives as an optional substance, and the meat is to be treated at a suitable temperature.

Literatures extending over many years points out that chicken and meat products are liable to be contaminated with various kinds of microorganisms from different sources and such contamination may render products unsafe to consumer or impair their utility especially in undeveloped countries where the hygienic measures are still underway [1-9].

The presence of Enterobacteriaceae, Coliforms and Staphylococci in meat and chicken products depend upon the meat used for grinding, sanitary conditions, practices during preparation, time and temperature of processing and storage. Also, during cutting and handling, meat surfaces exposed to ambient air provide excellent media for most bacteria.

*Escherichia coli* (*E. coli*) is a commensal microorganism whose niche is the mucous layer of the mammalian colon. This bacterium is the most abundant facultative anaerobe of the human intestinal microflora. Furthermore, *E. coli* is widely distributed in the intestinal tracts of warm blooded animals [2,10-17]. *E. coli* is often nonpathogenic, although different strains may cause diseases in gastrointestinal, urinary, or central nervous systems [3,18-27].

*Staphylococcus aureus* (*S. aureus*) is a major bacterial human pathogen that causes a wide variety of clinical manifestations. It is found in the environment and is also found in normal human flora, located on the skin and mucous membranes (most often the nasal area) of most healthy

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individuals. Also, it can be found in other areas of human contact including soil, water, and food products. *S. aureus* is considered the third worldwide cause amongst the food-borne illnesses reported cases [4,28-34].

*Salmonella* is a member of the *Enterobacteriaceae*, Gram negative, motile, with peritrichous flagella and non-spore forming rods. Also, *Salmonella* is a facultative anaerobic bacterium. However, *Salmonella* is not included in the group of organisms referred to as coliforms [5,35-43]. More than 2,500 different types of *Salmonella* exist, some of which cause illness in both animals and people. Some types cause illness in animals but not in people [6,44-52].

*Shigella* is Gram-negative, facultative anaerobic, non-spore forming, non-motile, rod-shaped bacteria closely related to *Salmonella*. Infection due to other *Shigella* species including *S. flexneri*, *S. boydii* and *S. dysenteriae* also occur. In contrast to the most common foodborne pathogenic agents, shigellosis is exclusively a human disease [7,53-60].

So the aim of the current study was to conduct a microbiological evaluation of chicken and beef sausages retailed for sale in different shops at the city of Al Ramadi, Iraq.

## Materials and Methods:

### Collection of samples:

A total of 100 of fresh chicken and beef sausage samples (50/each) were randomly collected from different shops at the city of Al Ramadi, Iraq. Each sample (250 g) was kept in a separate sterile polyethylene package and transferred directly with a minimum of delay to the laboratory in an insulating refrigerated container under possible aseptic condition to avoid any changes in the quality of the sample.

### Preparation of samples for microbiological examinations (APHA, 2001):

Samples were firstly cauterized by using hot spatula then cauterized parts were removed by using sterilized scalpel and forceps. Under aseptic conditions, 25 g of each sample were aseptically transferred into sterile blender flask containing 225 ml of sterile peptone water 1% and homogenized at 14000 rpm for 2.5 minutes. The mixture was left for 15 minutes at room temperature in order to achieve harmony. The contents of the flask were thoroughly mixed by shaking and 1 ml was transferred into a separate sterile tube containing 1 g of 0.1 % sterile peptone water from which tenth fold serial dilutions up to  $10^{-6}$  were prepared.

### Microbiological evaluation:

It was performed according to the guide lines recommended by ISO 4833, (2003) [8,61-70].

- **Determination of aerobic plate count:**

One ml of the previously prepared dilution was aseptically transferred into sterile Petri dish then about 10 ml of standard plate count agar previously melted and tempered at 45° C were added and thoroughly mixed in a horizontal position. After solidification, inoculated as well as control plates were incubated at an inverted position at 37° C for 48 hours. The colonies on the plates counted and the total aerobic bacterial count/gram was calculated.

- **Coliforms and *Escherichia coli* counts:**

From each of the previously prepared sterile dilutions, 0.1ml aliquots were delivered into duplicate sets of petri dishes, previously inoculated with 10ml of sterile violet red bile agar (VRb-agar), after sufficient spreading a cover layer of approximately 5 ml of (VRB-Agar) (tempered promptly to about 45° C) were poured over all plates. The plates were incubated at an inverted position at 37° C for 48 hours. All dark red colonies measuring 0.5 mm or more in diameter on uncrowded plates were then counted and the average number of colonies was calculated. From each similar colony from the plates, one colony was picked up and streaked on sterile Eosin Methylene blue (EMB) agar plates and incubated at 37° C for 24 hours then one colony was picked up from EMB plate into sterile semisolid agar tubes then the tubes were kept in refrigerator until further identification of *E. coli*.

- **Determination of *Staphylococcus aureus* count:**

The use of mannitol salt agar is highly recommended for enumeration of *Staphylococci*. From each of the serial dilutions previously prepared, 0.1

ml was inoculated onto the surfaces of separate plates and evenly spread with a sterile bent glass rod until the surface of the medium appears dry. Duplicate plates were prepared for each dilution. The plates were incubated at an inverted position at 35-37° C for 24-48 hours. Yellow colonies surrounded by yellow haloes were counted as mannitol fermenter. Biochemical tests as the blood plasma enzyme coagulation test, the catalase test, Gram straining, urea enzyme test and a group of other tests were performed upon them.

- **Determination of *Salmonella* and *Shigella* count:**

*Salmonella* and *Shigella* were isolated by using the selective medium Tetrathionate broth then 0.1 ml of pre-enriched samples were transferred to 10 ml Rappaport Vassiliadis broth (RV) and incubated at 42.5° C for 24 hours. Loopful from enriched RV broth was separately streaked onto Xylose Lysine Desoxycholate (XLD) agar and incubated at 37° C for 24 hours. Typical colonies were selected and streaked onto Triple sugar iron agar (TSI) and incubated at 37° C for 24 hours. Suspected colonies were subjected to biochemical identification using indole, methyl red, Voges Proskaur, citrate utilization and hydrogen sulphide production tests.

### Statistical Analysis:

Data collected were subjected to analysis of variance (ANOVA) by the aid of SAS, (2014) [9], then conducting T. test to assess the significance between different groups, and the level of significance is (P<0.05).

## Results:

**Table 1: Microbiological evaluation of chicken sausage**

Microbiological criteria	Minimum	Maximum	Mean	SD
Aerobic bacterial count	3.61×10 <sup>5</sup>	3.70×10 <sup>5</sup>	3.65×10 <sup>5</sup>	3.050
Coliforms count	4.7×10 <sup>4</sup>	6.0 ×10 <sup>4</sup>	5.35×10 <sup>4</sup>	4.031
<i>Escherichia coli</i>	1.5×10 <sup>3</sup>	2.2×10 <sup>3</sup>	1.85×10 <sup>3</sup>	2.426
<i>Staphylococcus aureus</i>	0.2×10 <sup>2</sup>	0.6 ×10 <sup>2</sup>	0.4×10 <sup>2</sup>	13.200
<i>Salmonella spp.</i>	0.02×10 <sup>2</sup>	0.06 ×10 <sup>2</sup>	0.04×10 <sup>2</sup>	1.418
<i>Shigella spp.</i>	0.0	0.01×10 <sup>2</sup>	0.005×10 <sup>2</sup>	0.418

SD means Standard Deviation

**Table 2: Microbiological evaluation of beef sausage**

Microbiological criteria	Minimum	Maximum	Mean	SD
Aerobic bacterial count	2.5×10 <sup>6</sup>	2.7×10 <sup>6</sup>	2.6×10 <sup>6</sup>	7.604
Coliforms count	1.7×10 <sup>6</sup>	2.2×10 <sup>6</sup>	1.9×10 <sup>6</sup>	14.449
<i>Escherichia coli</i>	2×10 <sup>3</sup>	3×10 <sup>3</sup>	2.5×10 <sup>3</sup>	3.910
<i>Staphylococcus aureus</i>	0.8×10 <sup>2</sup>	0.9×10 <sup>2</sup>	0.85×10 <sup>2</sup>	2.722
<i>Salmonella spp.</i>	0.08×10 <sup>2</sup>	0.09×10 <sup>2</sup>	0.085×10 <sup>2</sup>	0.495
<i>Shigella spp.</i>	0.01×10 <sup>2</sup>	0.02×10 <sup>2</sup>	0.015×10 <sup>2</sup>	0.485

**Table 3: Statistical analysis of mean values of microbiological criteria of chicken and beef sausage**

Microbiological criteria	Mean		T. test
	Chicken Sausage	Beef Sausage	
Aerobic bacterial count	3.65×10 <sup>5</sup>	2.6×10 <sup>6</sup>	89.535
Coliforms count	5.35×10 <sup>4</sup>	1.9×10 <sup>6</sup>	- 64.829
<i>Escherichia coli</i>	1.85×10 <sup>3</sup>	2.5×10 <sup>3</sup>	- 12.323
<i>Staphylococcus aureus</i>	0.4×10 <sup>2</sup>	0.85×10 <sup>2</sup>	- 27.785
<i>Salmonella spp.</i>	0.04×10 <sup>2</sup>	0.085×10 <sup>2</sup>	- 23.073
<i>Shigella spp.</i>	0.005×10 <sup>2</sup>	0.015×10 <sup>2</sup>	- 12.586

There were great differences according to T. tests and for each investigated microbiological criteria (P<0.01).

## Discussion:

In case of malpractices in handling, cooking or post-cooking storage of products, contamination of them with pathogens remains a crucial public health issue, leading to illness. Food production and health care cost increase is partially caused by food borne illness in developed countries which might also cause mortality, particularly in developing regions, where the health status of many individuals is already compromised.

Aerobic bacterial count is used as an indicator of bacterial population and it is based on an assumption that each cell will form a visible colony when examined with agar containing the appropriate nutrients. It is generic test for microorganisms that grow aerobically at mesophilic temperature but do not differentiate types of bacteria [10,71-75]. Also, it gives an idea about the hygienic measures applied during processing and also helps in the determination of the keeping quality of the poultry carcasses. So, it is the most reliable method for detection of sanitary level of proper processing, storage and marketing of food products.

The presented data in Table (1) showed that aerobic bacterial count for chicken sausages ranged from  $3.61 \times 10^5$  to  $3.70 \times 10^5$  with a mean value of  $3.65 \times 10^5$  cfu/g while the recorded results in Table (2) clarified that aerobic bacterial count for beef sausages ranged from  $2.5 \times 10^6$  to  $2.7 \times 10^6$  with a mean value of  $2.6 \times 10^6$  cfu/g. Moreover, it was clear that the mean value of aerobic bacterial count for both chicken and beef sausage exceeded the permissible limits (ABC must not exceed  $10^5$  cfu/g) meaning that samples were unfit for human consumption and reflecting a serious state of contamination during the processing of these sausages and probably contaminated meat and raw material besides inefficient cooking.

Coliforms are referred as general indicator microorganisms to measure the potential presence of enteric pathogens in foods, besides the measuring of fecal contamination of food products and the sanitary condition in the foods processing environment [11,76-82]. The presence of these organisms in food products depicts a deplorable state of poor hygiene and sanitary practices employed in the processing and packaging of this food product [12,83-90].

The presented data in Table (1) showed that coliforms count for chicken sausages ranged from  $4.7 \times 10^4$  to  $6.0 \times 10^4$  with a mean value of  $5.35 \times 10^4$  cfu/g while the recorded results in Table (2) clarified that coliforms count for beef sausages ranged from  $1.7 \times 10^6$  to  $2.2 \times 10^6$  with a mean value of  $1.9 \times 10^6$  cfu/g. Moreover, it was clear that the mean value of coliforms count for both chicken and beef sausage exceeded the permissible limits ( $10^5$  cfu/g) meaning that samples were unfit for human consumption. Unfortunately, undercooked meat products have caused much food poisoning incidence associated with coliforms and investigations had established that the bacteria are present in the feces and intestine contents that it could be potentially contaminate meat during the slaughtering process [13,91-96].

*E. coli* is considered the parameter of choice to evaluate the effectiveness of sanitation practices and potential fecal contamination of meat [14,97-100]. The presented data in Table (1) showed that *E. coli* count for chicken sausages ranged from  $1.5 \times 10^3$  to  $2.2 \times 10^3$  with a mean value of  $1.85 \times 10^3$  cfu/g while the recorded results in Table (2) clarified that coliforms count for beef sausages ranged from  $2 \times 10^3$  to  $3 \times 10^3$  with a mean value of  $2.5 \times 10^3$  cfu/g. The presence of *E. coli* in food may be due to handlers who contaminate food via manual contact or the respiratory tract by coughing and sneezing and also contamination may occur after heat treatment of the food [15].

In general, Staphylococci exist in air, dust, sewage and food or on food equipment, environmental surfaces, humans and animals. Humans are the primary reservoirs as they are present in the nasal passages and throats and on the hair and skin of 50 % or more of healthy individuals [16,101-103] therefore, the high count of staphylococci in food products indicates the presence of cross contamination, which is usually related to human skin, hand touch, discharge from human and clothing because of faulty handling activities, as they are typical contaminants from hands, clothes and utensils [17].

As shown in Table (1), *Staphylococcus aureus* count for chicken sausages ranged from  $0.2 \times 10^2$  to  $0.6 \times 10^2$  with a mean value of  $0.4 \times 10^2$  cfu/g while the recorded results in Table (2) clarified that aerobic bacterial count for beef sausages ranged from  $0.8 \times 10^2$  to  $0.9 \times 10^2$  with a mean value of  $0.85 \times 10^2$  cfu/g. Presence of *Staphylococcus aureus* in the examined samples made them non comply with the hygienic standards

and unfit for consumption as the thermal process used during manufacture can limit staphylococcal contamination but cannot eliminate preformed toxins [18].

Several Staphylococci food poisoning outbreaks were attributed to the use of bare hands in preparation of food [19]. There are many types of staphylococci, but most infections are caused by *Staphylococcus aureus* which is the third most common cause of confirmed food poisoning in the world [20].

CDC estimates *Salmonella* causes about 1.2 million illnesses, 23,000 hospitalizations, and 450 deaths in the United States every year. Food is the source for about 1 million of these illnesses. Most persons infected with *Salmonella* develop diarrhea, fever, and abdominal cramps 12 to 72 hours after infection. The illness usually lasts 4 to 7 days, and most persons recover without treatment. However, in some persons, the diarrhea may be so severe that the patient needs to be hospitalized [21].

The presented data in Table (1) showed that *Salmonella spp.* count for chicken sausages ranged from  $0.02 \times 10^2$  to  $0.06 \times 10^2$  with a mean value of  $0.04 \times 10^2$  cfu/g while the recorded results in Table (2) clarified that coliforms count for beef sausages ranged from  $0.08 \times 10^2$  to  $0.09 \times 10^2$  with a mean value of  $0.085 \times 10^2$  cfu/g.

The presence of *Salmonella* in sausages may reflect the unsatisfactory hygienic conditions and faults in personal hygiene during handling, packaging and marketing.

*Shigella* is one of the leading bacterial causes of diarrhea worldwide, causing an estimated 80–165 million cases. The number of deaths it causes each year is estimated at between 74,000 and 600,000. It is in the top four pathogens that cause moderate-to-severe diarrhea in African and South Asian children [22].

The presented data in Table (1) showed that *Shigella spp.* count for chicken sausages ranged from 0.0 to  $0.01 \times 10^2$  with a mean value of  $0.005 \times 10^2$  cfu/g while the recorded results in Table (2) clarified that coliforms count for beef sausages ranged from  $0.01 \times 10^2$  to  $0.02 \times 10^2$  with a mean value of  $0.015 \times 10^2$  cfu/g. The most common symptoms of *Shigella* infection are diarrhea, fever, nausea, vomiting, stomach cramps, and flatulence. It is also commonly known to cause large and painful bowel movements. The stool may contain blood, mucus, or pus. Hence, *Shigella* cells may cause dysentery [23].

The results of statistical analysis found in Table (3) showed a significant difference between the mean values of the studied microbiological criteria for chicken and beef sausages where the examined samples of beef sausages scored higher mean values than that of chicken sausages clarifying that beef sausages were highly contaminated with different types of bacteria reflecting bad hygienic conditions during processing and alarming the bells of dangers about the health issues of people consuming such products.

## Conclusion:

According to the recorded results in the current study, it was clear that both chicken and beef sausages retailed for sale in Al Ramadi were highly contaminated with different species of bacteria either from animal or human sources that would throw the light on manufacturing chain starting from animal farms passing by abattoirs then manufacture plants. Workers included in the manufacture process could be a significant source of contamination of sausages and they could be at the same time a significant barrier to prevent contamination if they received efficient training programs and the principles of Hazard Analysis and Critical Control Points (HACCP) and Good Manufacturing Practice (GMP).

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